Orphan Nuclear Receptor TR3/Nur77 is a Specific Therapeutic Target for Hepatic Cancers

Yingling Zeng1*, Xiaoguang Ye1, Degui Liao1, Shizhang Huang1, Huinan Mao1, Dezhang Zhao1,2 and Huiyan Zeng1,3

Abstract

Objective: Although great success has been achieved in cancer treatment, current cancer therapies, including anti-tumorigenesis and anti-angiogenesis, still face the problems of insufficient efficacy, resistance and intrinsic refractoriness, in addition to their toxic side effects. There is a demand to identify additional targets that can be blocked to turn off the downstream effects of most, if not all, pathways. Our previous studies suggest that orphan nuclear receptor TR3 (human)/Nur77 (mouse) is such a target. However, the correlation of TR3 expression and clinical tumor progression has not been studied.

Methods: The expression of TR3 was analysed in human primary hepatic carcinoma specimens from patients that have complete medical records with immunohistochemical staining. The statistical analysis was used to assess the significance of TR3 expression in tumor tissues, paratumor tissues and normal tissues, and to investigate the correlation of TR3 expression and clinicopathologic characteristics.

Results: TR3 is highly expressed in human hepatic cancer tissues, but not in normal liver tissues. The positive expression yields of TR3 are 67.67% (14/21), 19.05% (4/21) and 0% (0/10) in cancer tissues, para tumor tissues, and normal liver tissue, respectively, which are statistic significant (χ²=17.07, p<0.005). The expression of TR3 is significantly higher in cancer tissues than in para cancer tissues (χ²=9.722, p<0.005) and in normal tissues (p<0.0005). The levels of TR3 expression in human hepatic cancer tissues correlates well with tumors that are at low/middle degree of tumor differentiation and have portal vein thrombosis, metastasis and recurrence, but not with age, gender, tumor number and Alpha-fetal protein (AFP) volume.

Conclusion: The results indicate that TR3 is a specific therapeutic target for hepatic cancers.

Keywords

TR3; Nur77; Tumor; Hepatic cancer; Expression; Progression; Clinical; Correlation; Metastasis; Therapeutic target

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of Department of Pathology, the Second Affiliated Hospital of Guangzhou Medical University, P. R. China, between January 2010 to December 2013. Twenty-one human primary liver cancer specimens from patients that have complete medical records were retrieved. Approval of the current project was obtained from the local ethics committee.

The main characteristics of 21 patients were summarized in Table 1. Ages of all patients in this study are between 32 and 70 years (median, 58 years), with 13 cases of males and 8 cases of females. There are 8 cases and 13 cases with the tumor diameter less/equal or greater than 5 cm, respectively. Histological analysis indicated that 13 cases and 8 cases belong to low/middle and highly differentiated tumors, respectively. Among the 21 patients, 14 and 13 patients complicated with portal vein thrombosis and tumor metastasis, respectively.

Compliance with ethical standards

The research involved human samples. The study was approved by the Ethics Committee of The Second Affiliated Hospital of Guangzhou Medical University.

Tissue processing and Immunohistochemical staining

All tissues were fixed with 10% formalin, embedded with paraffin. Immunohistochemical staining on 4 µm sections was performed with an antibody against TR3/Nur77 (Univ-bio, Shanghai, CHINA), using immunological staining reagents following the protocol provided by the manufacture (Univ-bio, Shanghai, CHINA). PBS buffer was used as negative control.

Data analysis

Stained sections were analyzed by two independent pathologists. At least 10 fields with 400x amplification from each stained section were randomly chosen to be photographed with Olympus BX41 microscope (Olympus Scientific Solutions Americas Inc. Waltham, MA). Brown staining is referred as positive. Using half-quantitative measurement, the staining intensity was recorded as two levels, 1 as no staining and 2 as positive staining. The positive percentage of cancer cells was counted as 3 levels, level 1 as ≤ 25%, level 2 as 26%~75% and level 3 as ≥76%. The final score for each specimen was the multiplication of these two scores. They were referred to as high and ≥ 3 low and with <3, final score respectively. We found that the positive expression yields of TR3 are 67.67% (14/21), 19.05% (4/21) and 0% (0/10) in cancer tissues, paracancer tissues, and normal liver tissue, respectively, which are statistic significant (χ²=17.07, p<0.005). The expression of TR3 is significantly higher in cancer tissues than in paracancer tissues (χ²=9.722, p<0.005) and in normal tissues (p<0.0005) (Table 2). Further, TR3 is expressed in both cancer cells and vasculature of human hepatic cancer tissues, but not in normal liver tissues (Figure 1).

Correlation of TR3 expression and clinopathologic characteristics

The positive expression rate of TR3 is (a) 10/15 (66.7%) cases and 4/6 (66.67%) cases for age ≥ 60 and <60, respectively; and (b) 9/13 (69.23%) cases and 5/8 (62.5%) cases for males and females, respectively (Table 1). There is no significant difference of TR3 positive expression in terms of age and gender (p>0.5). Some, but not significant, difference of TR3 expression was found in 4/9 (44.44%) cases and 10/12 (83.33%) cases for tumors with a diameter ≤ 5 cm and >5 cm, respectively (Table 1, 0.05 < *p<0.5). Expression of TR3 is significantly higher in tumors with low/middle degree of differentiation, 11/13 (84.61%) cases, than in tumors with high degree of differentiation, 3/8 (37.5%) cases. Tumors with portal vein thrombosis showed significantly higher levels of TR3 expression, 12/14 (85.71%) cases, than tumors without portal vein thrombosis, 2/7 (28.57%) cases. TR3 is expressed in 11/13 (84.61%) cases tumors with metastasis, which is significantly greater than those tumors without

<table>
<thead>
<tr>
<th>Variables</th>
<th>No. of patients</th>
<th>Positive expression rate</th>
<th>p value</th>
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<td>Age (years)</td>
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<tr>
<td>≥ 60</td>
<td>15</td>
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</tr>
<tr>
<td>&lt; 60</td>
<td>6</td>
<td>4 (66.67%)</td>
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<td>Gender</td>
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<tr>
<td>Male</td>
<td>13</td>
<td>9 (69.23%)</td>
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<td>Female</td>
<td>8</td>
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<tr>
<td>Tumor diameter (cm)</td>
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<td></td>
<td></td>
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<tr>
<td>≤ 5</td>
<td>9</td>
<td>4 (44.44%)</td>
<td>0.080</td>
</tr>
<tr>
<td>&gt; 5</td>
<td>12</td>
<td>10 (83.33%)</td>
<td></td>
</tr>
<tr>
<td>Degree of tumor differentiation</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Low and middle</td>
<td>13</td>
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<td>0.040</td>
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<tr>
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<td>8</td>
<td>3 (37.5%)</td>
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<td>Portal vein thrombosis</td>
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<td></td>
<td></td>
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<tr>
<td>With</td>
<td>14</td>
<td>12 (85.71%)</td>
<td>0.017</td>
</tr>
<tr>
<td>Without</td>
<td>7</td>
<td>2 (28.57%)</td>
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<tr>
<td>Metastasis</td>
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<td></td>
<td></td>
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<tr>
<td>Yes</td>
<td>13</td>
<td>11 (84.62%)</td>
<td>0.040</td>
</tr>
<tr>
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<td>3 (37.5%)</td>
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<tr>
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<td>0.020</td>
</tr>
<tr>
<td>No</td>
<td>10</td>
<td>4 (40%)</td>
<td></td>
</tr>
<tr>
<td>Tumor number</td>
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<td></td>
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</tr>
<tr>
<td>Single</td>
<td>14</td>
<td>10 (71.43%)</td>
<td>0.428</td>
</tr>
<tr>
<td>Multiple</td>
<td>7</td>
<td>4 (57.14%)</td>
<td></td>
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<tr>
<td>AFP volume</td>
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<tr>
<td>&gt; 400 µg/L</td>
<td>12</td>
<td>9 (75%)</td>
<td>0.318</td>
</tr>
<tr>
<td>&lt; 400 µg/L</td>
<td>9</td>
<td>5 (55.56%)</td>
<td></td>
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</table>

Table 1: Correlation of TR3 expression and clinopathologic characteristics.

Results

Expression of TR3/Nur77 in human hepatic cancer tissues, paracancer tissues and normal tissues

In order to study whether TR3/Nur77 is a potential target for human tumors, we studied the correlation of TR3 expression with hepatic cancers. Human normal liver tissues and hepatic cancer tissues were immunostained with an antibody against TR3/Nur77. The stained sections were analysed by two independent pathologists. At least 10 fields with 400x amplification were randomly chosen from each stained section. Using half-quantitative measurement, the staining intensity was recorded as two levels, 1 as no staining and 2 as positive staining. The positive percentage of cancer cells was counted as 3 levels, level 1 as ≤ 25%, level 2 as 26%~75% and level 3 as ≥76%. The final score for each specimen was the multiplication of these scores. They were referred to as high and ≥ 3 low and with <3, final score respectively. We found that the positive expression yields of TR3 are 67.67% (14/21), 19.05% (4/21) and 0% (0/10) in cancer tissues, paracancer tissues, and normal liver tissue, respectively, which are statistic significant (χ²=17.07, p<0.005). The expression of TR3 is significantly higher in cancer tissues than in paracancer tissues (χ²=9.722, p<0.005) and in normal tissues (p<0.0005) (Table 2). Further, TR3 is expressed in both cancer cells and vasculature of human hepatic cancer tissues, but not in normal liver tissues (Figure 1).

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metastasis, 3/8 (37.5%) cases. TR3 expression was found significantly higher in tumors with recurrence, 10/11 (90.91%) cases, than in tumors without recurrence, 4/10 (40%) cases (Table 1, all \( \text{\textit{p}}<0.05 \)). Single tumors, 10/14 (71.43%) cases, did not show any difference in TR3 expression from multiple tumors, 4/10 (40%) cases. There is no difference of TR3 expression in tumors with AFP >400 µg/L, 9/12 (75%) cases, or <400 µg/L, 5/9 (55.56%) cases (Table 1, \( \text{\textit{p}}>0.5 \)). These data clearly indicate that expression of TR3 in human hepatic cancer tissues correlates very well with tumors that are at low/middle degree of tumor differentiation, have portal vein thrombosis, metastasis and recurrence, somehow with tumor diameter, but does not correlate with age, gender, tumor number and AFP volume.

Conclusions

The data presented here indicate that TR3 is highly expressed in human hepatic cancer tissues, but not in human normal liver tissues. The positive expression yield of TR3 is 67.67% and 19.05% in cancer tissues and paracancer tissues, respectively. Expression of TR3 in human hepatic cancer tissues correlates very well with tumors that are at low/middle degree of tumor differentiation and have portal vein thrombosis, metastasis and recurrence, but does not have any difference in regarding of age, gender, tumor number and Alpha-fetoprotein volume.

Angiogenesis is critical for tumor growth [15-19]. Anti-VEGF neutralizing antibodies and VEGFR kinase/multiple kinase inhibitors have been successfully developed and widely used in the clinic [20]. However, in addition to their toxic side effects [21] VEGF-targeted therapies in cancer face the problems of insufficient efficacy [22-31], resistance, and intrinsic refractoriness [29,32,33]. Current anti-angiogenesis therapies mainly focus on targeting a single molecule or a single pathway, although there are some combination therapies that target a couple or a few pathways. The map of signaling pathways resembles a spider web. Hence, targeting one or a few major pathways results in other pathways becoming dominant. It is desirable to identify additional angiogenesis targets, blocking of which will turn off the downstream effects of most, if not all, pathways. Our studies found that orphan nuclear receptor TR3 / Nur77 is a master transcription factor that is down-stream of almost, if not all, signaling pathways, including VEGF-A, histamine and serotonin that control the various transcription profiles in pathological angiogenesis [2-4]. Microvessel permeability induced by VEGF-A, histamine or serotonin, and tumor growth are almost completely inhibited in Nur77 knockout mice [2-4]. However, Nur77 null mice are viable, fertile, develop an apparently normal adult vasculature [5], and have no defects in normal skin wound healing whereas tumor growth is inhibited in Nur77-/- mice. These findings strongly support our hypothesis that TR3 is required for pathological angiogenesis.

TR3/Nur77 is induced by epidermal growth factor (EGF) and by serum in cancer cells, and knockdown of TR3/Nur77 inhibits the proliferation of tumor cells [34]. TR3/Nur77 plays important roles in cancer cell biology [9-12]. Our results that TR3 is expressed in both endothelial cells and tumor cells correlate very well with the function

<table>
<thead>
<tr>
<th>Tissues</th>
<th>No. of patients</th>
<th>TR3 expression level</th>
<th>( \chi ^2 )</th>
<th>( \text{\textit{p}} )</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>High (%)</td>
<td>Low or no (%)</td>
<td></td>
</tr>
<tr>
<td>Cancer tissues</td>
<td>21</td>
<td>14/21 (67%)</td>
<td>7/21 (33%)</td>
<td>17.07</td>
</tr>
<tr>
<td>Paracancer tissues</td>
<td>21</td>
<td>4/21 (19%)</td>
<td>17/21 (81%)</td>
<td></td>
</tr>
<tr>
<td>Normal tissues</td>
<td>10</td>
<td>0/10 (0%)</td>
<td>10/10 (100%)</td>
<td></td>
</tr>
</tbody>
</table>

Table 2: Expression of TR3 in human hepatic cancer tissues, paracancer tissues and normal tissues.
of TR3 in tumor cells. Targeting TR3 will inhibit not only tumor angiogenesis, but also tumorigenesis induced by EGF and other serum factors.

Other problems that VEGF-targeted therapy faces are resistance and intrinsic refractoriness that may be regulated by three mechanisms, 1) other angiogenic factors, such as hepatocyte growth factor (HGF) and fibroblast growth factor (FGF), regulate angiogenesis [35]; 2) tumor metastasis is increased [36]; 3) tumors increase resistance to anti-VEGF/VEGFR therapy [37]. The results obtained from current studies indicate that expression of TR3 is significantly increased in tumor tissues with metastasis and recurrence.

It was reported that TR3/Nur77 mediates VEGF-A-induced thrombomodulin expression to protect mice from arterial thrombus formation [38]. However, we found that TR3 expression is significantly increased in tumor tissues with portal vein thrombosis. The function of TR3 in mouse hepatic tumor model with or without thrombosis needs to be further studied.

In summary, we found that expression of TR3 correlates very well with the stages of hepatic cancers. To our knowledge, this is the first report about the clinical correlation of TR3 expression and human cancers. The results strongly support our previous hypothesis that TR3 is an excellent specific target for human cancers.

**Consent to Publish**

Informed consent was obtained from all individual participants included in the study.

**Authors’ Contributions**

YZ and HZ have the overall responsibility to the manuscript. They contributed to design all experiments, analyze and interpret data, write and approve the final version of manuscript to be published. XY participated in the design of experiments. DL acquired all data. SH performed analysis and interpretation of data. HM took part in analysis of Data. DZ took part in analysis and interpretation of data, and final approval of the version to be published.

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**References**


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