



Case Report

A Rare Case of Isodicentric Xq28 that Causes Mental Retardation: Molecular Characterization and Review

Gonzalez C^{1*}, Gutierrez M¹, Ruiz M², Huete B², Gonzalez S³, Gallego J⁴ and Cava F¹

Abstract

Study background: Isodicentric X chromosome is a rare structural abnormality in which the arms of the chromosome are mirror images of each other with two centromeres. Little description is available about characterization or phenotype-genotype association in isodicentric Xq28. While some of the females are normal, most only present premature ovarian failure or amenorrhea, but mental retardation is not usually reported.

Methods: We present a study of cytogenetic and molecular characterization of an isodicentric Xq28 found in a development retarded girl with dysmorphic features. G and C banding, CGH array and Bisulfite Methylation techniques were used.

Results: The study showed an almost total trisomy of X chromosome (Xper→Xq28) with a 1.10 Mb deletion in Xq28:qter, with two centromeres and a total skewed inactivation of one X chromosome. The deletion in Xq28 in this patient includes genes that are implicated in X-linked mental retardation and autism spectrum disorder.

Conclusion: Molecular studies help to elucidate genes that are implicated to understand the phenotype. The mental retardation and dysmorphic features in our patient can be attributed to the affected genes, their regulation and tissue expression. Although mental retardation is not usually reported this possibility has to be taken into account in order to suggest a genetic counselling in prenatal diagnosis and de novo findings. Literature on isodicentric X chromosomes with various breakpoints on Xq is reviewed and summarized in this report.

Keywords

Isodicentric chromosome; Cytogenetic; CGH-array; Mental retardation; Deletion; Duplication

Introduction

An isodicentric X chromosome (idic(X)), consists of duplicated portions of a single X chromosome with symmetrical banding patterns on opposite sides of the breakpoint and two centromeres [1]. Many mechanisms have been suggested for the formation of isodicentric X, and recently, some authors hypothesized that most idic(X)

chromosomes result from non-allelic homologous recombination between palindromic low copy repeats and long interspersed nuclear elements (LINE element) [2].

Sex chromosome abnormalities generally have less deleterious clinical effects as compared to autosomal alteration probably due to skewed inactivation. However, selective inactivation of the structurally rearranged X chromosome does not inevitably confer phenotypic normality in females and the presence of a complex rearranged X chromosome very often involves some phenotypic alterations.

The signs and symptoms vary based on the size and location of the duplication/deletion, the genes involved and the sex of the affected person. Males with X chromosome rearrangements are more severely affected than females and often show intellectual disability and sexual abnormalities [3]. In females, the phenotype remains unclear, lack of Xp may cause Turner syndrome with short stature due to haplo insufficiency of short stature homeobox-containing gene (SHOX), a gene that is involved in cell growth regulation [4]. Breakpoints distal to Xq25 are not related to any phenotypic anomalies apart from a few cases of secondary amenorrhea or premature menopause [5].

Duplications in Xp are rare, tall stature is described to be due to overexpression of SHOX [6]. However duplications of upstream and downstream of SHOX are also associated to Leri-Weill dyschondrosteosis [7]. Recently, microduplications in SHOX had also been related with neurodevelopment disorders [8].

Rearrangements with deletions in Xq may confer primary or secondary ovarian failure, in part as a result of loss of premature ovarian failure (POF) region of Xq27 associated with this phenotype [9]. Mental retardation is not described in these patients.

The present report describes a cytogenetic and molecular analysis of a dicentric X chromosome consisting two X chromosomes linked by the terminal Xq region. The karyotype was a non mosaic 46,X,idic(X)(q28) which consisted in an almost complete X trisomy with a loss of Xq28 region. The patient was a 9 years old girl referred for mild mental retardation, peculiar phenotype, behavioural disorders and toe-walking. The aim of this study was to identify the genetic background of the pathogenic phenotype in our patient and review of literature.

Case and Methods

A 9 years old girl was referred to the genetics laboratory to perform a karyotype. There was no family history of mental retardation, dysmorphic features or sexual dysgenesis.

On examination she was tall with height 1, 42 cm (p>99), her weight was 42 kg (p>99), she presented bradypsychia and stereotypes, round face with hypertelorism, broad nasal root, thick lips, obesity, and tiptoe walking.

Endocrine and metabolic studies were according to her Tanner stage II.

G banding was performed using standard techniques on metaphases. C-banding with Ba(OH)₂ and fluorescence in situ hybridization (FISH) technique with centromeric probe for X and Y chromosomes were carried out. Multiplex ligation-dependent probe

*Corresponding author: Cristina González, H.I. Sofia, Genetic Department, Sebastian de los Reyes Madrid, Spain, Tel: 0034914842324; E-mail: cgonzalez@brsalud.es

Received: April 18, 2017 Accepted: June 24, 2017 Published: June 30, 2017

amplification (MLPA) (SALSA MLPA PO70-B3, MRC-Holland) was performed for all telomeres (1p to 22p/Xp- 1q to 22q/Xq). CGH Array with 180000 probes platform (Agilent Technologies) and Bisulfite genomic method for X chromosome DNA methylation were performed.

Results

Cytogenetic studies

Karyotype analysis in 20 metaphases at 550-650 bands demonstrated 46 chromosomes with a normal X and restructured chromosome formed with two X chromosomes linked by the end of terminal q arm (Xq28) (Figure 1). Parental karyotypes were normal.

FISH technique with centromeric probe for X and Y chromosomes showed three signals for X chromosome in all nucleus, confirming single cell line with isodicentric X.

C-Banding confirmed the presence of two centromeres, Ba(OH)₂ observations showed one centromere darker than the other (Figure 2).

Molecular studies

MLPA technique showed deleted vesicle-associated membrane protein (VAMP) probe (located in Xq, PAR2 region,) and SHOX (PAR1) probe duplication. MLPA in parental blood samples was normal.

CGH array in patient assay showed an almost total trisomy of X chromosome (Xper->Xq28) with a 1.10 Mb deletion in Xq28:qter. This deletion includes 13 OMIM genes (Figure 3).

Coagulation factor VIII (F8), mature T-cell proliferation 1 (MTCP1), BRCA1-BRCA2-containing complex subunit 3 (BRCC3), von hippel-lindau binding protein 1 (VBP1), RAS-associated protein RAB39B (RAB39B), chloride intracellular channel 2 (CLIC2), H2A histone family, member b3 (H2AFB3), factor viii-associated gene 1 (F8A1), epsilon-trimethyllysine hydroxylase (TMLHE), sprouty RTK signaling antagonist 3 (SPRY3, VAMP7 and other non OMIM genes: FUN14 domain-containing protein 2 (FUNDC2), mature T-cell proliferation 1 neighbor (MTCP1NB), LOC100507404, H2A histone family member B1(H2AFB1), H2AFB1, H2A histone family member B2 (H2AFB2), LOC100507404. No patients with Xq28 deletions were described in the consulted data bases (Signature Genomics and Decipher).

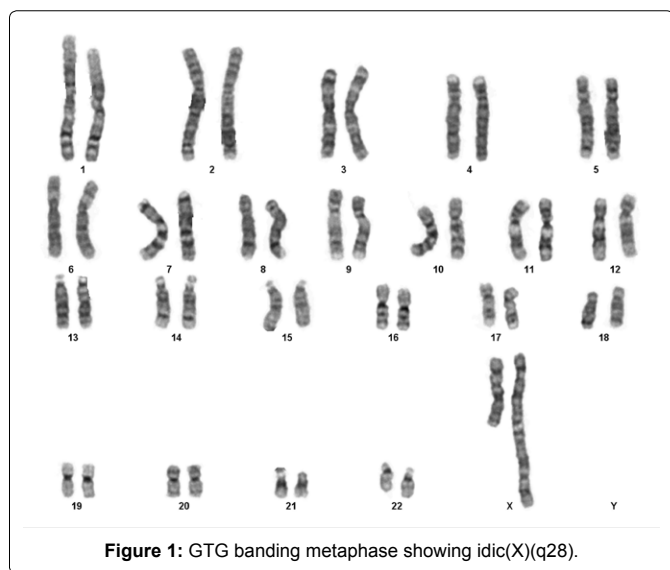


Figure 1: GTG banding metaphase showing idic(X)(q28).

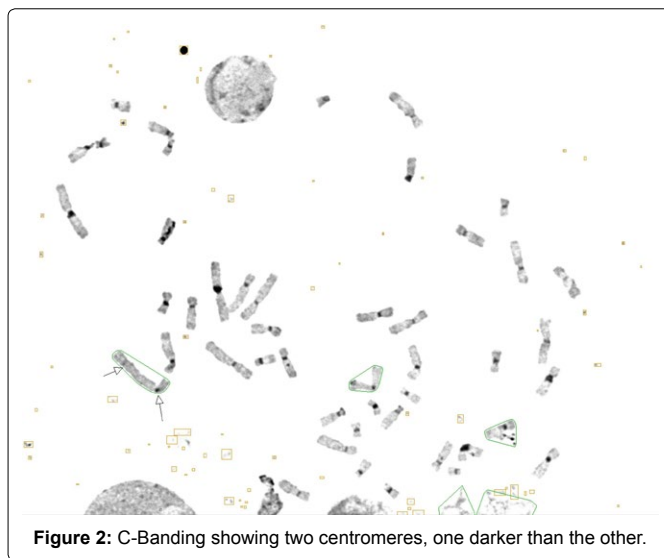


Figure 2: C-Banding showing two centromeres, one darker than the other.

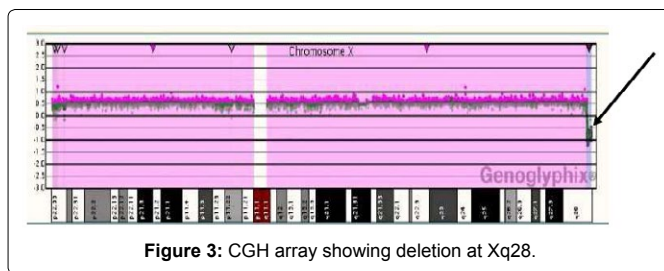


Figure 3: CGH array showing deletion at Xq28.

Bisulfite genomic method for DNA methylation showed a total skewed methylation of one chromosome X.

Discussion

Isodicentric X chromosomes fused by the short arms generally cause gonadal dysgenesis, short stature or Turner stigmata, while those attached by the long arms cause normal or taller than average stature, gonadal dysgenesis, amenorrhea, lack sexual development or infertility, but developmental delay is not usually reported [5].

Patients as the one we described idic(X)(q28) are very rare and the phenotype and structure is not well described in the literature. Similar isodicentric was long time ago described with secondary amenorrhea and normal intellectual development [10] while the patient we report showed mild mental retardation and dysmorphic features.

While patients with idic(X)(q28) are described with little sexual development our patient had already presented first sexual development signs at 9 years old. The critical region for gonadal dysgenesis is X, Xq13->q27 is conserved in our patient, and therefore the absence of gonadal dysgenesis could be expected. Premature ovarian failure or secondary amenorrhea might appear later as it is described in literature in patients with idic(X)(q28) [5].

Most of the i(X)q are isodicentric or pseudoisodicentric, because the activity from one of the centromeres is suppressed. The presence of two active centromeres could predispose to a mosaic line with 45,X and Turner stigmata due to the instability in the cellular division. Studies of dicentric chromosomes provide strong support for epigenetic mechanisms of centromere inactivation that include partial deletions of the alpha satellite array [11]. In plants, smaller centromeres are preferentially inactivated [12]. C-Banding in our

patient showed two distantly located centromeres, one centromere darker than the other, which could be indicating that the bigger is the active centromeres, which is consistent with the presence of a single line.

Preferential inactivation of the abnormal X is described in X rearrangement in peripheral blood [13]. However X inactivation in other tissues is difficult to demonstrate and could also play an important role in the clinical expression.

Partial duplications in SHOX gene are described in short, normal or tall stature patients leading to some uncertainty on the effect of SHOX duplication [14]. Duplication in our patient is not only limited to PAR1 but also to almost whole X chromosome. It is more likely that her tall stature is produced by the effect of X trisomy as in triple X syndrome. Microduplications at the SHOX locus are also described as low penetrance risk factor for autism and neurodevelopmental problems [8] as in the case here.

So far, no mental retardation is reported in patients with idic(X)(q28). However, this region is well known for its gene density [15]. Over 40 of approximately 300 X-linked diseases map to this region, molecular studies have led to the identification of causative genes for many diseases [16]. The deletion in Xq28 in this patient includes genes that are implicated in hemophilia, DNA damage response, X-linked mental retardation (RAB39B and CLIC2) and autism spectrum disorder (TMLHE, SPRY3). The highest expression of RAB39B is in neuronal precursors and neurons in the hippocampus. Mutation and deletions in RAB39B are described in patients with intellectual disability, autism and early onset Parkinson disease [17].

X-linked forms of intellectual disability are 3.5 times more common than autosomal forms. X-linked genes are highly expressed in the brain, and the proportion of X-linked genes expressed in the brain is significantly higher than that in other somatic tissues [18]. In females, phenotypic effects differ depending on whether the mutated gene is subjected or not subjected to X control inactivation [19]. Some genes escape X-inactivation and are expressed from both the active and inactive X chromosome, for instance it is unclear whether the phenotypic features of Klinefelter syndrome are caused by altered hormonal dosage or by the increased expression levels of escaped genes.

Short stature is correlated with proximal breakpoints while tall stature phenotype is correlated with distal breakpoint. Gonadal dysgenesis, POF, primary or secondary amenorrhea are also common problems of idic(X) when there is a 45,X line mosaicism, but this is not fulfilled in patients with a single line. There is not a specific correlation between breakpoint and all the features described in literature, probably due to the different deleted/duplicated genes, the tissue expression and X inactivation.

Our review concludes that genotype-phenotype correlation in non mosaic cases of idic(X) is more difficult than in those with 45,X line. Molecular studies can help to elucidate which genes are implicated in the rearrangement to try to understand the phenotype. The mental retardation and dysmorphic features in our patient would be produced by the affected genes, their regulation and tissue expression. Although mental retardation is not usually reported this possibility has to be taken into account to suggest a genetic counselling in prenatal diagnosis and de novo findings.

Acknowledgment

Thanks to Amelia Queipo, Vanesa Barea, Rebeca Moreno and Begonia Rodríguez for technical support. Thanks to Isabel Lorda for comments that improved the manuscript.

References

1. Pallister PD, Meisner LF, Elejalde BR, Francke U, Herrmann J, et al. (1977) The pallister mosaic syndrome. *Birth Defects Orig Artic Ser* 13: 103-110.
2. Teschler-Nicola M, Killian W (1981) Case report 72: mental retardation, unusual facial appearance, abnormal hair. *Synd Ident* 7: 6-7.
3. Bielanska MM, Khalifa MM, Duncan AM (1996) Pallister-Killian syndrome: a mild case diagnosed by fluorescence in situ hybridization. Review of the literature and expansion of the phenotype. *Am J Med Genet* 65: 104-108.
4. Leube B, Majewski F, Gebauer J, Royer-Pokora B (2003) Clinical, cytogenetic, and molecular observations in a patient with Pallister-Killian syndrome with an unusual karyotype. *Am J Med Genet Part A* 123: 296-300.
5. Reynolds JF, Daniel A, Kelly TE, Gollin SM, Stephan MJ, et al. (1987) Isochromosome 12p mosaicism (Pallister mosaic aneuploidy or Pallister-Killian syndrome): Report of 11 cases. *Am J Med Genet* 27: 257-274.
6. Schinzel A (1991) Tetrasomy 12p (Pallister-Killian syndrome). *J Med Genet* 28: 122.
7. Kwee ML, Barth PG, Arwert F, Madan K (1984) Mosaic tetrasomy 21 in a male child. *Clin Genet* 26: 150-155.
8. Lopes VE, Wyatt P (1985) Prenatal diagnosis of tetrasomy 21. *Prenat Diagn* 5: 233-235.
9. Abad DE, Gabarre JA, Izquierdo AM, López-Sánchez C, García-Martínez V et al. (2006) Pallister-Killian Syndrome Presenting With a Complex Congenital Heart Defect and Increased Nuchal Translucency. *J Ultrasound Med* 25: 1475-1480.
10. Dutly F, Balmer D, Baumer A, Binkert F, Schinzel A (1998) Isochromosomes 12p and 9p: parental origin and possible mechanisms of formation. *Eur J Human Genet* 6: 140-144.
11. de Ravel TJ, Keymolen K, van Assche E, Wittevronghel I, Moerman P, et al. (2004) Post-zygotic origin of isochromosome 12p. *Prenat Diagn* 24: 984-988.
12. Hodge JC, Hulshizer RL, Seger P, St Antoine A, Bair J, et al. (2012) Array CGH on unstimulated blood does not detect all cases of Pallister-Killian syndrome: A skin biopsy should remain the diagnostic gold standard. *Am J Med Genet Part A* 158: 669-673.
13. Polityko AD, Goncharova E, Shaminga L, Drozdovskaja N, Podleschuk L, et al. (2005) Pallister-Killian syndrome: rapid decrease of isochromosome 12p frequency during amniocyte subculturing. Conclusion for strategy of prenatal cytogenetic diagnostics. *J Histochem Cytochem* 53: 361-364.
14. Genevieve D, Cormier-Daire V, Sanlaville D, Faivre L, Gosset P, et al. (2003) Mild phenotype in a 15-year-old boy with Pallister-Killian syndrome. *Am J Med Genet Part A* 116: 90-93.
15. Gilgenkrantz S, Droulle P, Schweitzer M, Foliguet B, Chadeaux B et al. (1985) Mosaic tetrasomy 12p. *Clin Genet* 28: 495-502.
16. Chen CP, Chien SC (2010) Prenatal Sonographic Features of Pallister-Killian Syndrome. *J Med Ultrasound* 18: 43-53.
17. Mowery-Rushton PA, Stadler MP, Kochmar SJ, McPherson E, Surti U et al. (1997) The use of interphase FISH for prenatal diagnosis of Pallister-Killian syndrome. *Prenat Diagn* 17: 255-265.
18. Blancato J, Hunt M, George J, Katz J, Meck JM (1992) Prenatal diagnosis of tetrasomy 12p by in situ hybridization: Varying levels of mosaicism in different fetal tissues. *Prenat Diagn* 12: 979-983.
19. Wilson R, Harrison K, Clarke LA, Yong SL (1994) Tetrasomy 12p (Pallister-Killian syndrome): Ultrasound indicators and confirmation by interphase fish. *Prenat Diagnosis* 14: 787-792.

Author Affiliation

Top

¹H.I. Sofia, Genetic Department, Sebastian de los Reyes Madrid, Spain

²Department of Pediatrics, Av. 9 de Junio, 2, 28981, Parla, Madrid, Spain

³ICM lab, Rúa María Barbeito, 61, Lugo, Spain

⁴Jiménez Díaz Foundation, Department of Genetics, Av. Reyes Católicos, 2, 28040, Madrid, Spain