



Targeting Alarmin Cytokines in the treatment of severe Asthma

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Editorial

Asthma is a heterogeneous chronic airway disease with distinct phenotypes, characterized by different immunopathological pathways, clinical features, disease severity, physiology, and response to treatment. The established cytologic phenotypes of asthma include eosinophilic, neutrophilic, mixed granulocytic, and paucigranulocytic phenotypes [1]. Despite recent advances in the diagnosis and treatment of asthma, some phenotypes of asthma are difficult to treat, and still pose an exceptional health and pharmacoeconomic burden [2]. Approximately 3.6-10% of patients with asthma have severe refractory disease, which is uncontrolled despite treatment with high-dose inhaled corticosteroids (ICS), long acting β_2 -agonists (LABA), and/or leukotriene receptor antagonists (LTRA) [3,4]. A smaller proportion of patients unresponsive to high-dose ICS, respond favourably to interleukin (IL) antagonists, such as mepolizumab and reslizumab (anti-IL-5), benralizumab (anti-IL-5R α), and dupilumab (anti-IL-4R α). T helper type (Th2) targeted monoclonal antibodies (mAb) are highly effective in the treatment of eosinophilic asthma and have a good safety profile [5-8]. They reduce exacerbation, improve lung function (forced expired volume in 1 sec., FEV₁), and the quality of life, and are steroid sparing [5-8]. However, they are mostly ineffective in patients with biomarker specific eosinophilic asthma, and have little therapeutic value in patients with other phenotypes of asthma, such as Th17-driven neutrophilic asthma, and paucigranulocytic asthma [9]. Biologics also do not ameliorate airway remodeling due to goblet cell hyperplasia and oversecretion of mucus, subepithelial basement thickening and fibrosis, airway smooth muscle (ASM) cell hyperplasia and hypertrophy [10-14]. Moreover, anti-Th2 biologic therapies decrease exacerbation rates by 48-59%.

There are no biologics targeting the Th17/IL-17 axis which play a pivotal role in the pathogenesis of neutrophilic asthma, one of the phenotypes of asthma unresponsive to biologics [9]. Busse et al. [15] in a randomized, placebo-controlled phase IIa trial of brodalumab, a monoclonal antibody against IL-17 receptor (IL-17R α), in patients with moderate-to-severe asthma, reported no statistically significant benefit in terms of Asthma Control Questionnaire-6 (ACQ-6) scores, FEV₁, or use of rescue short-acting β -agonists (SABA). O'Byrne and colleagues [16] in a larger, multicenter, dose finding trial, investigated AZD5069, a

selective prostaglandin D₂ receptor CXCR2 antagonist, in patients with uncontrolled asthma receiving medium-to-high dose ICS, and LABA. The findings in this clinical trial showed no significant benefits in the rates of exacerbations, ACQ-5, and FEV₁. Risankizumab a humanised IgG monoclonal antibody that targets the p19 subunit of IL-23 is in phase 2 clinical trials for the treatment of asthma [17]. However, blockade of IL-17 and IL-17R is more effective in the treatment of other chronic autoimmune diseases, such as psoriasis vulgaris, erythrodermic psoriasis, psoriatic arthritis, and inflammatory bowel disease [17]. Brodalumab (Siliq[®]) [18], secukinumab (Cosentyx[®]) [19], and risankizumab (Skyrizi[®]) [20] have been approved in several countries, and are excellent drugs for the treatment of plaque psoriasis.

Airway epithelium constitutes the first line of defense against allergens, chemicals, pollutants, and microbes, such as viruses, bacteria, and fungal spores, and trauma in the atmospheric environment [21,22]. This can lead to epithelial injury, and impaired epithelial barrier function. Dysfunctional allergic epithelium in response to allergens, pollutants, and viral respiratory infections release three bioactive cytokines nicknamed "alarmins", including IL-25, IL-33, and thymic stromal lymphopoietin (TSLP) [23-27]. The three "whistle blower" cytokines, although they belong to different cytokine families, play synergistic roles in the pathophysiology of severe asthma [27]. They stimulate Th2, innate lymphoid group 2 cells (ILC2), mast cells, basophils, and eosinophils to release large quantities of cytokines (IL-4, IL-5, IL-13, IL-25, IL-33, TSLP), chemokines (eotaxins 1-3, CXCL8, CXCL9, CXCL10, CXCL11, CXCL12, CCL17, CCL22), and growth factors (TGF- β 1, EGF-1, FGF, SCF, VEGF, angiogenin) responsible for airway hyperresponsiveness (AHR), and structural changes in the airways. Alarmin cytokines together with the pro-fibrotic Th2 cytokines IL-4 and IL-13 orchestrate airway remodeling, characterized by mucus hypersecretion, subepithelial fibrosis, increase in ASM mass and contractility, and progressive persistent airflow obstruction [28-30]. IL-25, IL-33, and TSLP are favourable targets for the development of new biologics for asthma treatment, and for prophylaxis of asthma, particularly due to asthma exacerbations provoked by respiratory viral infections [31].

Currently, there is no anti-IL-25, and anti-33 biologics approved for the treatment of asthma, including the eosinophilic phenotype. Tezepelumab a first-in-class monoclonal antibody that blocks TSLP is approved by the U.S. Food and Drug Administration (FDA) for the treatment of severe asthma without an eosinophilic phenotype in patients 18 years and older [32].

Thymic stromal lymphopoietin is a member of the 4-helix bundle cytokine, most closely related to IL-7 [33]. TSLP signaling pathway is mediated through its complex heterodimeric receptor formed by a TSLP-specific TSLPR subunit (CRLF2) and the IL-7 α signaling chain [34,35]. TSLP activates dendritic cells in response to allergen exposure, inducing naive CD4⁺ T cell differentiation to Th2 cells, which produce and secrete cytokines (IL-4, IL-5, IL-9, and IL-13), chemokines, and growth factors [36,37]. This leads to switching of B cells to produce IgE, degranulation of mast cells, and eosinophils, and airway eosinophilia. TSLP can also activate innate lymphoid group 2 cells (ILC2) to produce large amounts of IL-5, and IL-13 which drives Th2 immune responses [38]. There is

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evidence that TSLP also plays a central role in the pathogenesis of Th17-driven neutrophilic asthma. TSLP promotes dendritic cells to induce polarization of naïve CD4+ T cells into Th17 cells which produce IL-17 a master cytokine responsible for neutrophilic asthma [39,40].

TSLP contributes to airway remodeling by stimulating ASM cell proliferation and migration [41,42], and facilitates crosstalk between ASM cells and mast cells [43]. This leads to secretion of more pro-inflammatory cytokines and chemokines by both cell types [44-46]. Additionally, TSLP stimulates fibroblasts to produce collagen which promotes reticular basement membrane fibrosis [47,48]. Thus TSLP plays a critical role in airway remodeling and fixed airflow limitation.

TSLP seems attractive to target in therapeutic interventions to treat asthma because it is an upstream cytokine at epithelial barrier, and it promotes both eosinophilic and neutrophilic asthma, and paucigranulocytic asthma through its effects on airway remodeling [49].

Phase 1b [50], Phase 2b PATHWAY [51], and Phase 3 [52] clinical trials have documented the efficacy, safety profile of tezepelumab in the treatment of in patients with uncontrolled asthma. The first Phase 1b clinical trial evaluated the efficacy of tezepelumab in an allergen challenge model of asthma in patients with mild, allergic asthma [50]. Tezepelumab 200 mg administered intravenously every 4 weeks for 3 months resulted in a decrease in blood eosinophil count at 2 weeks of treatment, and the level of fractional exhaled nitric oxide (FeNO) improved after the first dose of tezepelumab. Bronchoprovocation with allergen at day 42 and 84 showed that tezepelumab treatment significantly inhibited the early and late asthmatic responses [50]. Phase 2b PATHWAY, multicentre, randomized, parallel-group, double-blind, placebo-controlled trial assessed the efficacy of tezepelumab as add-on therapy in patients with moderate-to-severe asthma and a history of exacerbations, and uncontrolled disease [51]. This dose-ranging study demonstrated that tezepelumab significantly reduced exacerbation rates by 62% (70 mg every 4 weeks), 71% (210 mg every 4 weeks), and 66% (280 mg every 4 weeks); respectively compared with placebo. There was also improvement in lung function, and biomarkers of eosinophilic asthma (blood eosinophil count and FeNO) in all the three treated groups. The improvements were observed in all the phenotypes of asthma, and independent of baseline blood eosinophil counts, IgE levels, and FeNO concentration [51]. This indicates that tezepelumab is effective in most phenotypes of asthma, which gives it an advantage [53]. The Phase 3 follow-up analysis of data of pro-inflammatory biomarkers and proteomics for patients who received tezepelumab 210 mg every 4 weeks showed that tezepelumab decreased serum IL-5 and IL-13 by 30% at 1 year, FeNO by 25%, and total serum IgE by 20% [54]. This indicates that tezepelumab is an effective biotherapeutic in the treatment of eosinophilic asthma.

Most recently, Gauvreau et al. [54] have shown that CSJ117, a fully human neutralizing antibody antigen-binding fragment (Fab) that belongs to IgG1 isotype subclass attenuated both the early asthmatic response (EAR), and late asthmatic response (LAR) in 28 patients with mild, atopic asthma at day 84 of the study. CSJ117 was associated with significantly higher minimum FEV1 during the LAR on day 84 ($P=0.038$), and the percentage decrease and maximum decrease in FEV1 from before the allergen inhalation challenge were significantly less in the CSJ117 group compared with placebo (4.20% vs 11.38%, $P=0.008$, and 9.28% vs 17.70%, $P=0.29$); respectively [54]. Similarly, during the EAR, both measures significantly less in the CSJ117 treated group compared with placebo. CSJ probably might be the first inhaled biologic for add on treatment for patients with asthma. The aforementioned study was also associated with a reduction in FeNO, a biomarker of IL-13-driven eosinophilic inflammation, and none

of the patient's withdrawal from the study due to adverse events.

In summary, biologics targeting the Th2-driven eosinophilic asthma are well established treatment for severe steroid unresponsive asthma, and have been shown to be safe and very effective in the treatment of severe uncontrolled disease. Tezepelumab an anti-TSLP antagonist has been demonstrated to be effective in the treatment of patients with both eosinophilic and neutrophilic asthma. The new kid in the biologics armament CSJ117 may prove to be effective and well tolerated as add on treatment for patients with mild asthma.

Reference

1. Simpson JL, Scott R, Boyle MJ, Gibson PG (2006) Inflammatory subtypes in asthma: Assessment and identification using induced sputum. *Respirology* 11(1): 54-61.
2. O'Neill S, Sweeney J, Patterson CC (2015) The cost of treating severe refractory asthma in the UK: An economical analysis from the British Thoracic Society Difficult Asthma Registry. *Thorax* 70:376-378.
3. Moore WC, Meyer DA, Wenzel SE, Teague WG, Li H, et al. (2010) Identification of asthma phenotypes using cluster analysis in the Severe Asthma Research Program. *Am J Respir Crit Care Med* 181:315-323.
4. Chung KF, Wenzel SE, Brozek JL, Bush A, Castro M, et al. (2014) International ERS/ATS guidelines on definition, evaluation and treatment of severe asthma. *Eur Respir J* 43:343-373
5. Pavord ID, Korn S, Howarth P, Bleecker ER, Buhl L, et al. (2012) Mepolizumab for severe asthma (DREAM): A multicenter, double-blind, placebo-controlled trial. *Lancet* 380:651-659.
6. Castro M, Zangrilli J, Wechsler ME, Bateman ED, Brusselle GG, et al. (2015) Reslizumab for inadequately controlled asthma with elevated eosinophil count: Results from two multicentre, parallel, double-blind, randomised, placebo-controlled, phase 3 trials. *Lancet Respir Med* 3:355-366
7. Busse E, Bleecker R, FitzGerald JM, Ferguson GT, Barker P, et al. (2019) Long-term safety and efficacy of benralizumab in patients with severe, uncontrolled asthma: 1-year results from the BORA phase 3 extension trial. *Lancet Respir Med* 7:46-59.
8. Castro M, Corren J, Pavord ID, Maspero J, Wenzel S, et al. (2018) Dupilumab efficacy and safety in moderate-to-severe uncontrolled asthma. *N Engl J Med* 378: 2486-2496.
9. Syabbalo N (2020) Clinical features and management of neutrophilic asthma. *J Pulm Med Respir Res*; 6:036.
10. Ebina M, Takahashi T, Chiba T (1993) Cellular hypertrophy and hyperplasia of airway smooth muscles underlying bronchial asthma. A 3-D morphometric study. *Am Rev Respir Dis* 148:720-726.
11. Niimi A, Matsumoto H, Amitani R, Nakano Y, Mishima M, et al. (2000) T. Airway wall thickness in asthma assessed by computed tomography. Relation to clinical indices. *Am J Respir Crit Care Med* 162(4 Pt 1):1518-1523.
12. Munakata M (2006) Airway remodeling and airway smooth muscle in asthma. *Allergol Int* 55:235-245.
13. Girodet PO, Ozier A, Triani T (2011) The pivotal role of airway smooth muscle in asthma pathophysiology. *J Allergy* 742710

14. Ozier A, Allard B, Bara I, Girodet PO, Trian T, et al. (2011) The pivotal role of airway smooth muscle in asthma pathophysiology. *J Allergy ID*: 742710.
15. Busse WW, Holgate S, Kerwin E, Chon Y, Feng J, et al. (2013) Randomized, double-blind, placebo-controlled study of brodalumab, a humanized anti-IL-17 receptor monoclonal antibody, in moderate to severe asthma. *Am J Respir Crit Care Med* 188(11):1294-1302.
16. O'Byrne PM, Metev H, Puu M, Richter K, Keen C, et al. (2016) Efficacy and safety of a CXCR2 antagonist, AZD5059, in patients with uncontrolled persistent asthma: a randomized, double-blind, placebo-controlled trial. *Lancet Respir Med* 4(10):797-806.
17. Moro LR (2018) Anti-IL-23p19 monoclonal antibody, Treatment of psoriasis, Treatment of bowel disease, Treatment of asthma. *Drugs of the Future* 43(5):331.
18. Foulkes AC, Warren RB (2019) Brodalumab in psoriasis: Evidence to date and clinical potential. *Drug Context* 8:212570.
19. Yang EJ, Beck KM, Liao W. (2018) Secukinumab in the treatment of psoriasis: patient selection and perspectives. *Psoriasis* 8:75-82.
20. McKeage K, Diggan S (2019) Risankizumab: First global approval. *Drugs* 79(8):893-900.
21. Heijink IH, Nawijn MC, Hackett TL (2014) Airway epithelial barrier function regulates the pathogenesis of allergic asthma. *Clin Exp Allergy* 44(5):620-630.
22. Hammad H, Lambrecht BN (2015) Barrier epithelial cell and control of type 2 immunity. *Immunity* 43:29-40.
23. Holgate ST (2007) Epithelial dysfunction in asthma. *J Allergy Clin Immunol*; 120(6):1233-1244.
24. Haworth O, Levy BD (2007) Endogenous lipid mediators in the resolution of airway inflammation. *Eur Respir J* ; 30(5):980-992.
25. Bartemes KR, Kita H (2012) Dynamic role of epithelium-derived cytokines in asthma. *Clin Immunol* 143(3):222-235.
26. Ziegler SF, Roan F, Bell BD, Stoklasek TA, Kitajima M, et al. (2013) The biology of thymic stromal lymphopoietin (TSLP). *Adv Pharmacol* 66:129-155.
27. Cayrol C, Girard JP (2014) IL-33: An alarmin cytokine with crucial roles in innate immunity, inflammation and allergy 31:31-37.
28. Zhu Z, Homer RJ, Wang Z, Chen Q, Qeba GP, et al (1999) Pulmonary expression of interleukin-13 causes inflammation, mucus hypersecretion, subepithelial fibrosis, physiologic abnormalities, and eotaxin production. *J Clin Invest* 103:779-788.
29. Hough KP, Curtiss ML, Blain TJ, Liu R-M, Trevo J, et al. (2020) Airway remodeling in asthma. *Front Med*.
30. Syabbalo N (2020) Clinical features and management of eosinophilic asthma. *J Respir Dis Treat* 1:105.
31. Beale A, Jayaranaman A, Jackson DJ, McIntyre JD, Edwards MR, et al. (2014) Rhinovirus-induced IL-25 in asthma exacerbation drives type 2 immune allergic pulmonary inflammation. *Sci Transl Med* 2014; 6:256.
32. Tezpeumab Granted Breakthrough Therapy Designation by US FDA for the treatment of patients with severe asthma without an eosinophilic phenotype [news release]. Amgen's website. Accessed September 10, 2018.
33. Leonard WJ (2002) TSLP: finally in the limelight. *Nat Immunol* 3:605-607.
34. Park LS, Martin U, Garka K, Glinak B, Di Santo JP, et al. (2000) Cloning of the murine thymic stromal lymphopoietin (TSLP) receptor. Formation of a functional heterometric complex require interleukin 7 receptor. *J Exp Med* 192:659-670.
35. Tonozuka Y, Fujio K, Sugiyama T, Nosaka T, Hirai M, et al. (2001) Molecular cloning of a human novel type 1 cytokine receptor related to 1/TSLPR. *Cytogenet Cell Genet* 93:23-25.
36. Watson B, Gauvreau GM (2014) Thymic stromal lymphopoietin: A central regulator of allergic asthma. *Expert Opin Ther Targets* 18(7):771-785.
37. Zhang Y, Zhou B (2012) Functions of thymic stromal lymphopoietin in immunity and disease. *Immunol Res* 52(3):211-223.
38. Klose CS, Artis D (2016) Innate lymphoid cells as regulators of immunity, inflammation and tissue homeostasis. *Nat Immunol* 17(7):765-774.
39. Tanaka J, Watanabe N, Kido M, Saga K, Akamatsu T, et al. (2009) Human TSLP and TLR3 ligands promote differentiation of Th17 cells with a central memory phenotype under Th2-polarized conditions. *Clin Exp Allergy* 39(1):89-100.
40. Gao H, Ying S, Dai Y (2017) Pathological roles of neutrophil-mediated inflammation in asthma and its potential for therapy as target. *J Immunol Res* 3743048.
41. Fehrenbach H, Wagner C, Wegmann M (2017) Airway remodeling in asthma: what really matters. *Cell Tissue Res* 367(3):551-569.
42. Redhu NS, Shan L, Movassagh H, Gounni AS (2013) Thymic stromal lymphopoietin induces migration in airway smooth muscle cells. *Sci Rep* 3:2301.
43. Kaur D, Doe C, Woodman L, Heidi Wan WY, Sutcliffe A, et al. (2012) Mast cell-airway smooth muscle crosstalk: the role of thymic stromal lymphopoietin. *Chest* 142(1):76-85.
44. Damera G, Tliba O, Panettieri RA (2009) Airway smooth muscle as an immunomodulatory cell. *Pulm Pharmacol Ther* 22(5):353-359.
45. Berger P, Girodet, Begueret H (2003) Tryptase-stimulated human airway smooth muscle cells induce cytokine synthesis and mast cell chemotaxis. *FASEB J* 17:2139-2141.
46. Amin K, Janson C, Boman G, Venge P (2005) The extracellular deposition of mast cell products is increased in hypertrophic airway smooth muscle in allergic asthma but not in nonallergic asthma. *Allergy*; 60:12411247.
47. Cao L, Liu F, Liu T, Wu J, Zhao J, et al. (2018) TSLP promotes asthmatic airway remodeling via p38-STAT3 signaling pathway in human fibroblast. *Exp Lung Res* 44(6):288-301.
48. Wu J, Liu F, Zhao J, Wei J, Lv J, et al. (2013) Thymic stromal lymphopoietin promotes asthmatic airway remodelling in human lung fibroblast cells through STAT3 signalling pathway. *Cell Biochem Funct* 31(6):496-503.
49. Syabbalo N (2020) Clinical features and management of

- paucigranulocytic asthma. *Ann Clin Med Res* 1(3):1011.
50. Gauvreau GM, O'Byrne PM, Boulet LP, Wang Y, Cockcroft D, et al. (2014) Effect of anti-TSLP antibody on allergen-induced asthmatic responses. *N Engl J Med* 370(22):2102-2110.
 51. Corren J, Parnes J, Wang L, Mo M, Roseti SL, et al. (2017) Tezepelumab in adults with uncontrolled asthma. *N E J Med* 377(10):936-946.
 52. Pham TH, Ren P, Parnes JR, Griffiths JM (2019) Tezepelumab reduces multiple key inflammatory biomarkers in patients with severe uncontrolled asthma in the Phase 2b PATHWAY study. *Am J Respir Crit Care Med*; 119:A2679.
 53. Marone G, Spadaro G, Braile M, Poto R, Criscuolo G, et al. (2019) Tezepelumab: A novel biological therapy for treatment of severe uncontrolled asthma. *Expert Opin Investig Drugs*; 28(11):931-940.
 54. Gauvreau GM, Hohlfeld JM, Grants S, Jain M, Cabanski M, et al. (2020) Efficacy and safety of an inhaled anti-TSLP fragment in adults with mild atopic asthma. *Am J Respir Crit Care Med* 201:A4207.

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