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Short Communication

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Relationship between Cellular Senescence and Redox Potential on Adult T-Cell Leukemia Cells

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Abstract

Human T-cell leukemia virus type I (HTLV-I) is a human retrovirus and an etiologic agent of adult T-cell leukemia/lymphoma (ATL/ ATLL). The subtypes acute and lymphoma have a poor prognosis with median survival rate of approximately 6 months. Some excellent therapeutic strategies for ATL are required. Retinoids [all-*trans* retinoic acid (ATRA) and tamibarotene (Am-80) have been reported to inhibit the *in vitro* growth of HTLV-1 [+] T-cell lines and that of fresh cells obtained from patients with adult T-cell leukemia (ATL). We showed a clinical efficacy of retinoid therapy in treating ATL has also been evaluated. Further, we found that retinoid induced cellular senescence and redox potential on ATL cells.

Keywords

HTLV-I; Retinoid; Senescence; ROS; Redox

Introduction

Human T cell leukemia virus type I (HTLV-I) is a human retrovirus that is an etiologic agent of adult T cell leukemia/lymphoma (ATL/ATLL) [1]. We previously reported the clinical efficacy of alltrans retinoic acid (ATRA) for patients with adult-T-cell leukemia (ATL) [2]. Additionally, we found that retinoids including ATRA could facilitate cellular senescence in HTLV-I-positive T-cell lines and fresh primary cells obtained from patients with acute ATL [3]. Cellular senescence was detected by staining for senescence-associated β -galactosidase (SA β -Gal) [4]. It has known that senescence is a well-established anti-cancer mechanism [5] and can be caused by telomere attrition, DNA damage, oncogene and oxidative stress [6].

In our previous study, significant cellular senescence was observed for *Tax* mRNA expressing cells (HUT102, MT-2, MT-4, and ATL-2 cells) than ED40515 cells, which do not express *Tax* mRNA because of a nonsense mutation [3]. Thus, we assume that *Tax* is an oncogene that could possibly be involved in oncogene-induced senescence (OIS) by retinoids. It has been reported that OIS does occur in human and mouse tumor cells *in vivo* [7,8]. Furthermore, it is caused by accumulation of DNA damage, which in turn causes oncogene-driven

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accumulation of reactive oxygen species (ROS) [9]. Our hypothesis is that the *Tax* oncogene may induce cellular senescence via ROS, but the number of these senescing cells is easily reduced by retinoid. These results indicated that Tax mRNA expression may be a critical factor in retinoid therapy for patients with ATL and retinoid may be a reasonable agent for ATL with facilitating cellular senescence.

The balance of oxidative/anti-oxidative influences may play an important role in the modulation of cellular function. It has been reported that L-cysteine and L-cystine act as a buffer of the redox potential of the environment in cells or serum [10,11]. In our previous study, to study the effects of exogenous thiol compounds on the sensitivity to retinoid in a HTLV-I positive T cell line, ATL2 cells [12] were cultured with thiol compounds (L-cystine, GSH and TRX), following addition of ATRA or 13-cis RA [13,14]. Preincubation of ATL-2 with L-cystine or GSH resulted in complete restoration of growth despite the inhibitory effects of RA; this suggested that it helped to increase the redox potential of the intracellular environment [10]. These processes are antagonized by antioxidants such as cysteine and GSH [11]. Moreover, the effects of thiol compound, N-acetyl cysteine (NAC) for cellular senescence was examined. Figure 1 showed that spontaneous senescence was induced on HTLV-I positive T-cell lines (HUT102 and MT-2) but not on HTLV-I negative T-cell lines (Jurkat and MOLT-4). Furthermore, ATRA treatment induced more senescence on just HTLV-I positive T-cell lines. However, pretreatment with NAC for 24 hrs blocked cellular senescence by following addition with ATRA. These results suggest that the imbalance of intracellular redox potential in HTLV-I [+] T cell lines may be associated strongly with the sensitivity to RA and exogenous thiol compounds may prepare the intracellular environment to become resistant to RA. Further, pre-treatment with NAC for those cells was sufficient to recover from cellular senescence and finally rescue cell proliferation on HTLV-I positive T-lymphocytes. Having obtained those results described above, it has been suggested tight relationship between cellular senescence and redox potential on HTLV-I positive cells.

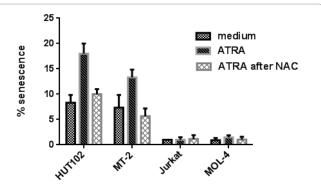


Figure 1: The effects of NAC for cellular senescence.

Note: Both HTLV-I positive cell lines [HUT102 and MT-2] and HTLV-I negative T-cell lines [Jurkat and MOLT-4] with 10-5 M NAC for 24 hrs were pre-incubated, and then 10-5 M ATRA was added to those cells for 24 hrs. Cellular senescence was detected by staining for senescence-associated β -galactosidase [SA β -Gal]. Values for the calculation were the mean of triplicate culture.

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